

Ultrafine Pd Nanoparticles Encapsulated in Microporous Co₃O₄ Hollow Nanospheres for In Situ Molecular Detection of Living Cells

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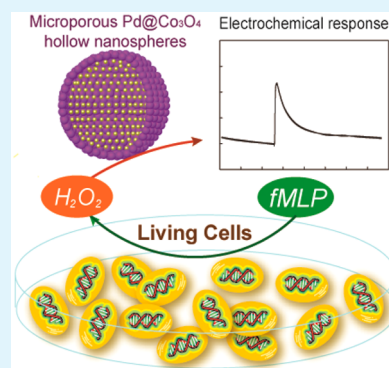
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Supporting Information

ABSTRACT: Recent progress in the in situ molecular detection of living cells has attracted tremendous research interests due to its great significance in biochemical, physiological, and pathological investigation. Especially for the electrochemical detection of hydrogen peroxide (H₂O₂) released by living cells, the highly efficient and cost-effective electrocatalysts are highly desirable. In this work, we develop a novel type of microporous Co₃O₄ hollow nanospheres containing encapsulated Pd nanoparticles (Pd@Co₃O₄). Owing to the synergy effect between the permeable microporous Co₃O₄ shell and the ultrafine Pd nanoparticles that encapsulated in it, the resultant Pd@Co₃O₄ based electrode exhibits excellent electrochemical sensor performance toward H₂O₂, even when the content of Pd in Pd@Co₃O₄ hollow nanospheres is as low as 1.14 wt %, which enable it be used for real-time tracking of the secretion of H₂O₂ in different types of living human cells.

KEYWORDS: microporous hollow nanospheres, ultrafine Pd nanoparticles, electrochemistry, biosensors, living cell tracking



1. INTRODUCTION

Reactive oxygen species (ROS), including superoxide anion (O₂^{•-}), hydrogen peroxide (H₂O₂), hydroxyl radical (OH[•]), and peroxynitrite (ONOO⁻), have attracted great attention due to their important roles in the metabolism process.^{1,2} H₂O₂ is one of the most stable ROS and the product of O₂^{•-} dismutation from superoxide dismutase. The physiological levels of H₂O₂ are closely associated with the degradation and formation of reactive free radicals, such as O₂^{•-} and •OH. Excessive H₂O₂ is implicated in arteriosclerosis, cancer, stroke, diabetes, and a host of other serious diseases.³ Therefore, it is of great importance to monitor the H₂O₂ level in a biological environment, especially in a cellular environment. Nowadays, several analytical techniques have been developed for the determination of H₂O₂, such as fluorometry,^{4,5} spectrophotometry,⁶ chromatography,⁷ and electrochemistry.⁸ Among these techniques, electrochemical methods are more suitable for in situ and real-time analysis of H₂O₂ due to their fast response, high sensitivity, good selectivity, facile operation, and excellent reproducibility.⁸ Electrochemical methods are usually based on the catalytic reduction of H₂O₂ by the natural enzyme, for example, horseradish peroxidase, which shows high catalytic efficiency and selectivity toward H₂O₂;^{9–11} however, the immobilization and stabilization protocol of the enzyme on the electrode is very complicated. Moreover, the activity of the enzyme electrode is easy to reduce or even deactivate.¹² In this case, noble metals (Pt, Pd, Ag, and Au) and their alloys have been explored as the substitute of enzyme for H₂O₂ detection due to their high catalytic activity toward the direct electro-

chemical oxidation/reduction of H₂O₂.¹³ Especially, tremendous efforts have been stimulated to the design and synthesis of Pd nanoparticles (NPs) owing to their size- and shape-dependent electrocatalytic performance,^{14–16} which gives rise to a significant improvement for nonenzymatic electrochemical sensing of H₂O₂.¹⁷ Nevertheless, in the design and application of these metal NP catalysts, the technologically crucial issue is the undesirable coalescence, sintering, aggregation, or corrosion/dissolution of NPs, which causes a rapid decay in catalytic abilities.^{18–20} On the other hand, for electrocatalytic applications, it is necessary to load noble metal NPs on an electrode with high density in order to achieve large active surface areas and high catalytic activity, which would inevitably increase the cost of the metal catalyst. Therefore, it is still a challenge to develop low-load noble metal NPs without sacrificing the high catalytic activity, in view of improving the efficiency and cost-effectiveness of a catalytic process.

To overcome these challenges, in this work, we develop a novel type of Pd-based nanocatalyst, i.e., microporous Co₃O₄ hollow nanospheres containing encapsulated Pd NPs (Pd@Co₃O₄), and explored its practical application as the electrode material in an electrochemical sensor for the nonenzymatic detection of H₂O₂ released by living cells. Previous works have also demonstrated that loading Pd nanoparticles on a large-surface-area and porous substrate is an effective way to enhance

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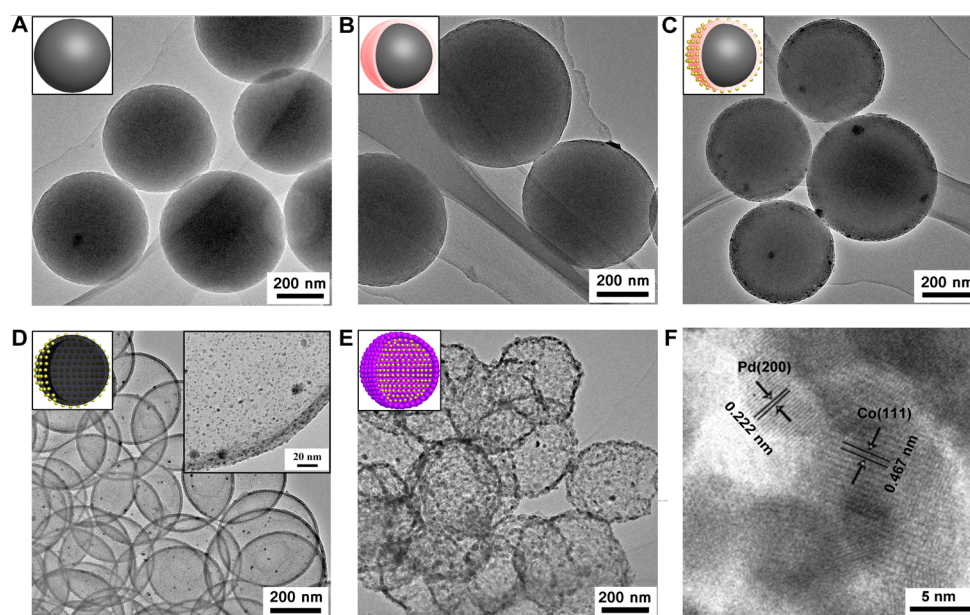


Figure 1. TEM images of (A) SiO₂ spheres, (B) SiO₂@PDA spheres, (C) SiO₂@PDA@Pd spheres, (D) C@Pd hollow spheres, and (E) Pd@Co₃O₄ hollow spheres and (F) HRTEM image of Pd@Co₃O₄ hollow spheres.

the electrocatalytic activity and improve the nonenzymatic sensing performance toward the H₂O₂.²¹ Recently, significant efforts have been devoted to fabricating hollow spheres with hierarchical core–shell structure owing to their multifunctional use in a myriad of fields, including surface enhanced Raman scattering (SERS), photocatalysis, drug delivery, Li-ion battery, and nanoreactor.^{22–33} For electrocatalytic application, the core–shell structured materials can effectively prevent the encapsulated catalytic active species from aggregation or leaching by the shell protection.³⁴ Meanwhile, the hollow nanospheres with microporous shells provide a large surface area for catalytic reaction as well as promote diffusion and mass transfer of reactants, which allow better reaction kinetics at the electrocatalyst surface. Owing to the synergy effect between the permeable microporous Co₃O₄ shell and the active Pd NPs encapsulated in it, the resultant Pd@Co₃O₄ based electrochemical sensor shows significantly improved electrocatalytic activity toward H₂O₂ as well as high sensitivity, even when the content of Pd in Pd@Co₃O₄ hollow nanospheres is as low as 1.14 wt %. These outstanding features, combined with its excellent stability and reproducibility, enable it to be used for real-time tracking of the secretion of H₂O₂ in living cells, i.e., human cervical carcinoma Hela cell, RSC96 Schwann cells, and human umbilical vein endothelial cells (HUVEC). These allow the Pd@Co₃O₄ based biosensor to contribute to the full realization of a high-performance biosensor for physiological and pathological monitoring of H₂O₂.

2. EXPERIMENTAL SECTION

2.1. Materials. All solutions were prepared using deionized water (resistivity >18 Ω·cm⁻¹). *N*-Formyl-methionyl-leucyl-phenylalanine (fMLP) and SiO₂ spheres with sizes ranging from 350 to 400 nm were purchased from Sigma-Aldrich. K₂PdCl₄ (99%) was purchased from Sinopharm Chemical Reagent Co. (China). Dopamine hydrochloride (98%) and tris(hydroxymethyl) aminomethane (Tris, 99.8%) were purchased from Aladdin Chemistry Co., Ltd., China.

2.2. Synthesis of Pd@Co₃O₄ Hollow Spheres. For the synthesis of SiO₂@PDA spheres, 1000 mg of the SiO₂ (350–400 nm) was immersed in 100 mL of 3 mg/mL dopamine Tris solution (pH 8.5, 10

mM Tris buffer) and allowed to proceed for 48 h under stirring at room temperature.³⁵ The resultant product was separated and collected and subsequently put through five wash cycles and dried by freeze-drying. A 900 mg portion of as-synthesized SiO₂@PDA was dispersed in 75 mL of DI water; then, 70 mg of K₂PdCl₄ was added, and the mixture was kept in a vial under vigorous stirring for 30 min in an ice bath.³⁶ The mixture was stirred at 0 °C for 30 min. After the reaction, the resulting Pd-NP-deposited SiO₂@PDA was separated from the suspension, and subsequently washed with ultrapure water five times and dried by freeze-drying. The content of Pd NPs in SiO₂@PDA@Pd can be adjusted by changing the amount of precursor K₂PdCl₄ that was added in the SiO₂@PDA aqueous dispersion. The resulting SiO₂@PDA@Pd composites were carbonized at 500 °C for 3 h under an inert atmosphere. Finally, SiO₂ cores were etched by HF solution (≈4%) to get C@Pd hollow spheres.³⁷ Then, C@Pd (20 mg) was dispersed in toluene (1.6 mL). Co₂(CO)₈ (12.4 mg) in toluene (6 mL) was added. The reaction mixture was heated at 100 °C overnight. During this process, the dark solution became transparent. The resultant dark yellow solids were retrieved by centrifugation, washed with toluene, and dried under a vacuum for 3 h. Then, the solids were heated at 500 °C for 5 h under air to form black powders,²⁶ and the Pd@Co₃O₄ hollow spheres were obtained. The Co₃O₄ hollow spheres were prepared under the same procedure but without the addition of K₂PdCl₄ in the SiO₂@PDA dispersion.

2.3. Fabrication of a Hollow Nanospheres Modified Electrode. The as-obtained Pd@Co₃O₄ powdery sample was dispersed in deionized water with ultrasonic agitating for a few minutes to give a 1.0 mg mL⁻¹ black suspension. Then, 10 μL of the suspension was dropped on a cleaned glass carbon electrode (GCE) and the solvent was evaporated in air; thus, a uniform film coated electrode (Pd@Co₃O₄/GCE) was obtained. The C@Pd/GCE was fabricated through a similar method. The as-prepared Pd@Co₃O₄/GCE and C@Pd/GCE were washed carefully with deionized water and then dried at room temperature. All the experiments were processed at room temperature. Before the electrochemical measurement, the nanohybrid electrodes were activated by cyclic voltammetric (CV) scanning for 20 cycles in the potential range from -0.4 to +1.0 V versus saturated calomel electrode (SCE).

2.4. Cell Culture and Detection of Extracellular Release of H₂O₂. In this work, three types of living human cells, i.e., human cervical carcinoma Hela cell, RSC96 Schwann cells, and human umbilical vein endothelial cells (HUVEC), were obtained from Center for Tissue Engineering and Regenerative Medicine, Union Hospital,

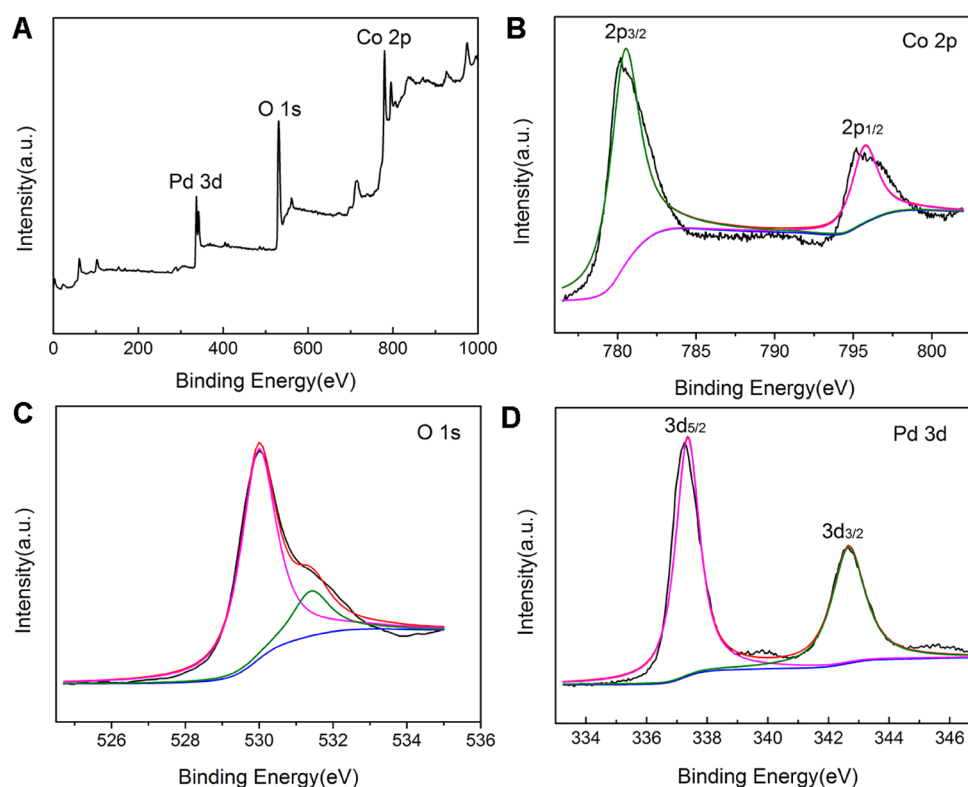


Figure 2. (A) XPS survey spectrum of Pd@Co₃O₄ hollow spheres. Curve fit of (B) Co 2p, (C) O 1s, and (D) Pd 3d spectra of Pd@Co₃O₄ hollow spheres.

HUST (Wuhan, China). The cells were maintained in a culture medium consisting of Dulbecco's modified Eagle medium (DMEM) at 37 °C and subcultured every 3 days. After centrifuging, the cells were planted at a confluency of 80% and used for the electrochemical experiments. To detect the secretion of H₂O₂ by living cells, Pd@Co₃O₄/GCE was used as the working electrode and located near the living cell lines in a culture medium consisting of DMEM at 37 °C. The well containing no living cells was used as the control. For electrochemical experiments, 10 μL of *N*-formyl-methionyl-leucyl-phenylalanine (fMLP, 0.1 mM) was injected into the testing wells and control wells, and the amperometric current responses at the applied potential of 0.6 V were recorded.

2.5. Characterization. CV and chronoamperometric experiments were performed with a CHI660E electrochemical workstation (CH Instrument Company, Shanghai, China). A conventional three-electrode system was adopted. The working electrode was a modified electrode, and the auxiliary and reference electrodes were Pt foil and SCE, respectively. Transmission electron microscopy (TEM) and high-resolution transmission electron microscopy (HRTEM) images were obtained using a TECNAI G2 20 U-Twin instrument (Netherlands) operated at an acceleration voltage of 200 kV. The samples were suspended in ethanol and were prepared by being drop-cast onto a carbon-coated 200-mesh copper grid and subsequently dried at room temperature. X-ray photoelectron spectroscopy (XPS) was performed with an ESCALAB MKII spectrometer (VG Co., U.K.), using Mg K α radiation (1253.6 eV) at a pressure of 2.0×10^{-10} mbar. The peak positions were internally referenced to the C 1s peak at 284.6 eV. The Pd content in the sample was determined using a microwave plasma-atom emission spectrometer (MP-AES, Agilent 4100, USA). The fluorescence microscope (Olympus IX71, Japan) equipped with a DP73 camera was used to examine sericin scaffolds under the light with the different wavelengths. The images were taken with the software cellSens standard 1.7 (Olympus, Japan).

3. RESULTS AND DISCUSSION

For the synthesis of Pd@Co₃O₄, the SiO₂ spheres with a good monodispersity and a uniform diameter of 350–400 nm (Figure 1A and Figure S1, Supporting Information) were first immersed in 3 mg/mL dopamine (DA) Tris solution for 48 h. During this procedure, DA will self-polymerize to form a transparent polydopamine (PDA) layer that wrapped on SiO₂ spheres (SiO₂@PDA), as shown in the transmission electron microscopy (TEM) of Figure 1B. Previous studies have demonstrated that PDA contained a certain amount of catechol groups, which are able to release electrons when oxidized into the corresponding quinone group and trigger reduction processes of metallic cations.^{38,39} Consequently, it can be used as a reducing agent to prepare PDA–NP composites via direct redox reaction with metal salt under the mild condition. In this work, K₂PdCl₄ was added into the SiO₂@PDA suspension under vigorous stirring for 30 min at 0 °C, upon which the spontaneous redox reaction between K₂PdCl₄ and PDA occurs, leading to the formation of ultrafine Pd NPs, which were fairly well-distributed on the surface of the SiO₂@PDA sphere (Figure 1C). The as-obtained Pd NPs decorated SiO₂@PDA (SiO₂@PDA@Pd) spheres were carbonized at 500 °C for 3 h under an inert atmosphere. Then, the SiO₂ cores were etched by HF solution (~4%) to form unique C@Pd hollow spheres, which was confirmed by TEM image (Figure 1D) and scanning electron microscopy (SEM) image (Figure S2, Supporting Information). From the high-resolution TEM (HRTEM) image, it can be observed that numerous ultrafine Pd NPs are dispersed on the surface of hollow spheres, with a mean size of ~2 nm (Figure 1D, inset). The successful synthesis of C@Pd was further confirmed by the energy dispersive X-ray spectroscopy (EDX), which demonstrates the

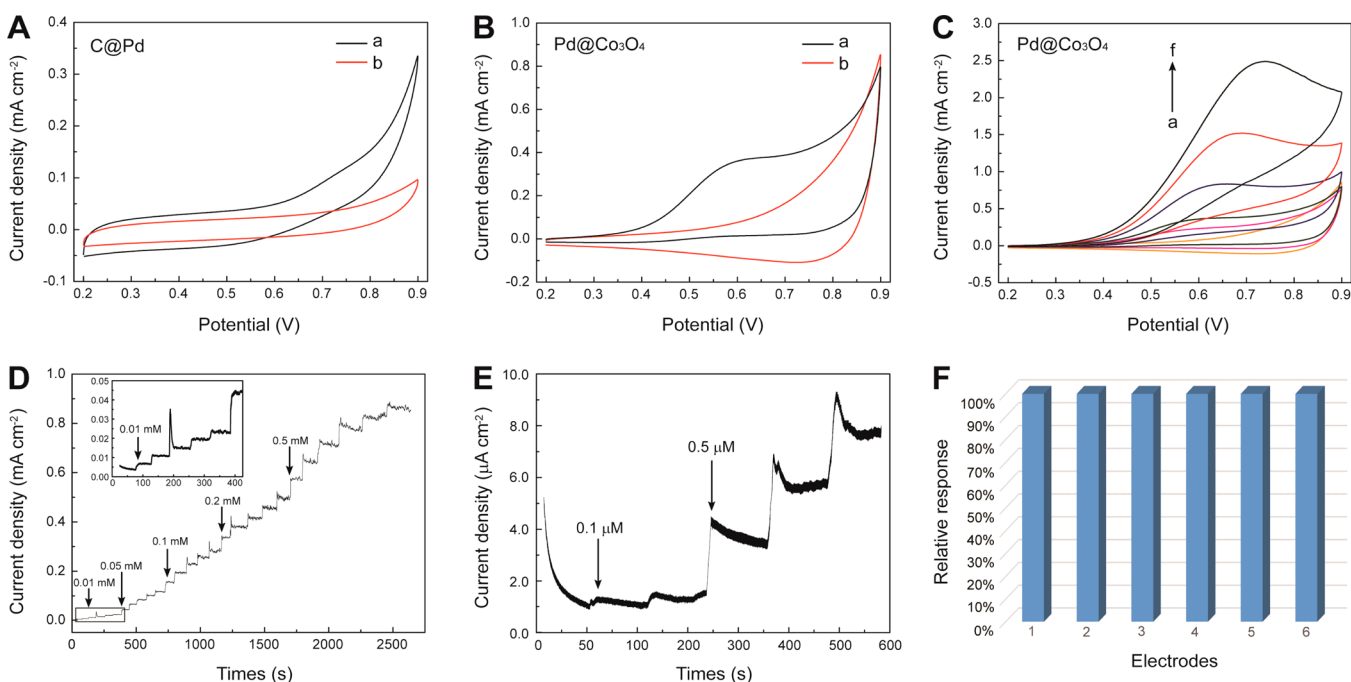


Figure 3. CV curves of (A) C@Pd/GCE and (B) Pd@Co₃O₄/GCE in 0.1 M PBS (pH 7.4) in the presence (A) and absence (B) of H₂O₂ (2 mM). (C) CV curves of Pd@Co₃O₄/GCE in 0.1 M PBS (pH 7.4) in the presence of H₂O₂ with different concentrations (from a to f: 0, 1, 2, 5, 10, 20 mM). Scan rate: 50 mV s⁻¹. (D) Typical amperometric response of Pd@Co₃O₄/GCE to successive addition of 0.01, 0.05, 0.1, 0.2, and 0.5 mM H₂O₂ in stirring PBS buffer (pH 7.4). The inset shows the partial enlarged curves. (E) Amperometric response of Pd@Co₃O₄/GCE to successive addition of 0.1 and 0.5 μM H₂O₂ in stirring PBS buffer (pH 7.4). (F) Amperometric response of six different Pd@Co₃O₄/GCEs to 0.1 mM H₂O₂. Applied potential: 0.6 V.

presence of C and Pd elements in the hollow spheres (Figure S3, Supporting Information). The content of Pd in C@Pd hollow nanospheres has been determined to be 2.08 wt % using MP-AES. The C@Pd hollow spheres were redispersed in toluene. After addition of Co₂(CO)₈ in C@Pd suspension, the reaction mixture was heated at 100 °C overnight, and the dark solution became transparent. The resultant dark yellow solids were retrieved by centrifugation, washed with toluene, and dried under a vacuum for 3 h. Then, the solids were heated at 500 °C for 5 h under air to form black powders. In this process, the C@Pd hollow spheres were converted to Pd@Co₃O₄ hollow spheres, which also exhibit a good monodispersity and a uniform diameter of 350–400 nm (Figure S4, Supporting Information). The content of Pd in Pd@Co₃O₄ hollow nanospheres is as low as 1.14 wt % using MP-AES measurement. Moreover, it can be observed that the Pd@Co₃O₄ hollow spheres contain microporous shells (Figure 1E and Figure S5, Supporting Information). Figure 1F shows the HRTEM image of Pd@Co₃O₄ hollow spheres, which indicates that the microporous shells actually consist of many tiny NPs. These NPs have a single crystalline structure extending along the Co(111) direction, with a lattice spacing of 0.467 nm.⁴⁰ And the single crystalline Pd(200) NPs with a lattice spacing of 0.222 nm are encapsulated in the Co₃O₄ microporous shells.⁴¹ The XRD pattern further demonstrates that Pd@Co₃O₄ hollow spheres exhibit six diffraction peaks corresponding to the (111), (220), (311), (400), (422), and (440) facets of cubic Co₃O₄ (JCPDS 74-2120). However, owing to the ultrafine size (~2 nm) and low content (1.14 wt %) of Pd NP Pd@Co₃O₄ hollow spheres, their diffraction peaks are undetectable in the XRD patterns recorded in the 2θ range of 10–90° (Figure S6, Supporting Information).

The surface chemical compositions and the valence states of samples are further investigated by the X-ray photoelectron spectroscopy (XPS). As shown in Figure 2A, the XPS spectrum of the Pd@Co₃O₄ sample exhibits three main peaks centered at the Co, O, and Pd core level regions, which are assigned to Co 2p, O 1s, and Pd 3d, respectively. The deconvoluted Co 2p XPS spectrum shown in Figure 2B exhibits two characteristic broad peaks encompassing the Co²⁺/Co³⁺ 2p_{3/2} and Co²⁺/Co³⁺ 2p_{1/2} states at 780.4 and 795.7 eV, respectively. The deconvoluted peaks centered at 530.0 and 531.4 eV can be observed in Figure 2C, which correspond to the oxygenated groups. These results combined suggest the formation of the standard Co₃O₄ phase.²⁷ The deconvoluted Pd 3d XPS spectrum exhibits two distinct peaks at 337.4 and 342.7 eV, respectively (Figure 2D), which are in good agreement with the reported XPS data of Pd 3d_{5/2} and Pd 3d_{3/2} in PdO.⁴²

To evaluate the electrochemical sensing performance of Pd@Co₃O₄ nanospheres, the powdery Pd@Co₃O₄ sample was dispersed in deionized water and modified on the clean glass carbon electrode (GCE) to fabricate the Pd@Co₃O₄ modified GCE (Pd@Co₃O₄/GCE). For comparison, the C@Pd/GCE was also prepared under the same procedure. Figure 3A shows the cyclic voltammetric (CV) behaviors of C@Pd/GCE in the absence and presence of 2.0 mM H₂O₂ in 0.1 M phosphate buffer solution (PBS) at a scan rate of 50 mV s⁻¹. In blank PBS (pH 7.4), the CV curve of Pd@Co₃O₄/GCE does not display any visible peaks in the potential range from 0.2 to 0.9 V. In the presence of 2 mM H₂O₂, a well-defined anodic peak that is associated with the oxidation of H₂O₂ can be observed at 0.58 V on Pd@Co₃O₄/GCE (Figure 3B). Along with increased concentration of H₂O₂ from 1.0 to 20 mM, the oxidation peak currents also increase dramatically (Figure 3C). In contrast, C@Pd/GCE gives rise to an inconspicuous oxidation peak at

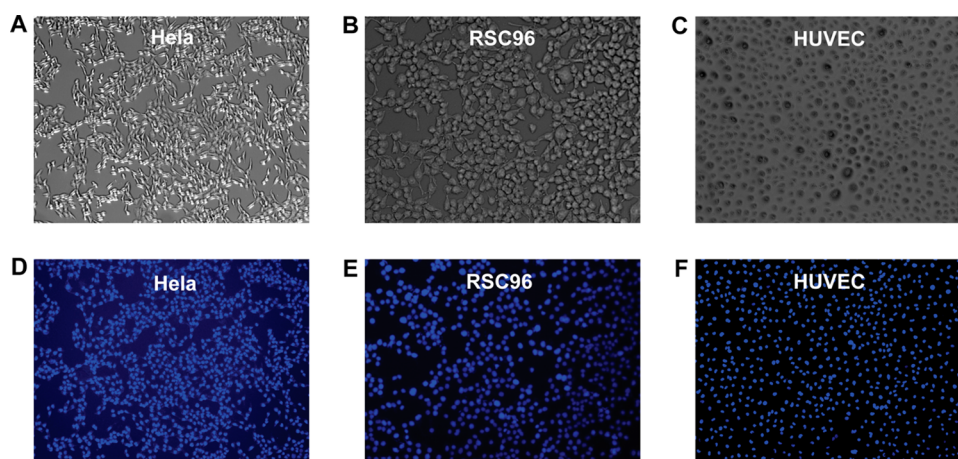


Figure 4. Bright-field image (A–C) and dark-field image (D–F) of HeLa cell, RSC96 Schwann cells, and HUVEC cells used in the in vitro tests.

0.75 V in 0.1 M PBS containing 2.0 mM H_2O_2 , which is 130 mV more positive than that of $\text{Pd@Co}_3\text{O}_4/\text{GCE}$. However, under the same condition, there are not any obvious redox peaks of H_2O_2 observed at $\text{Co}_3\text{O}_4/\text{GCE}$, in comparison with those without H_2O_2 . These indicate that the $\text{Pd@Co}_3\text{O}_4/\text{GCE}$ possesses high electrocatalytic activity toward H_2O_2 oxidation by taking advantage of the high electrocatalytic activity of Pd NPs on Co_3O_4 hollow spheres, as well as the synergistic effect between Pd NPs and Co_3O_4 hollow spheres, even when the content of Pd in $\text{Pd@Co}_3\text{O}_4$ hollow nanospheres is as low as 1.14 wt %.

Furthermore, the relationship between the electrocatalytic activity of $\text{Pd@Co}_3\text{O}_4$ hollow nanospheres toward the oxidation of H_2O_2 and Pd mass content in $\text{Pd@Co}_3\text{O}_4$ hollow nanospheres has also been investigated. For the preparation of $\text{Pd@Co}_3\text{O}_4$ hollow nanospheres, the Pd mass content in $\text{Pd@Co}_3\text{O}_4$ nanocomposites can be originally adjusted by changing the amount of precursor K_2PdCl_4 that is added in $\text{SiO}_2@\text{PDA}$ aqueous dispersion. The results show that the amperometric current responses of $\text{Pd@Co}_3\text{O}_4/\text{GCE}$ toward 2.0 mM H_2O_2 first increase with precursor K_2PdCl_4 content. However, when the mass ratio of K_2PdCl_4 to $\text{SiO}_2@\text{PDA}$ reaches 7/90, the amperometric responses tend to decrease (Figure S7, Supporting Information), probably due to the aggregation of Pd NPs on Co_3O_4 hollow nanospheres, which decreases the electrocatalytic activity of $\text{Pd@Co}_3\text{O}_4$ hollow nanospheres. Therefore, in this work, the optimal mass ratio of K_2PdCl_4 to $\text{SiO}_2@\text{PDA}$ is fixed at 7/90 for the further amperometric measure of H_2O_2 .

Figure 3D presents the amperometric responses of $\text{Pd@Co}_3\text{O}_4/\text{GCE}$ for successive addition of H_2O_2 under an applied potential at 0.6 V in 0.1 M PBS (pH 7.4). Upon each addition of H_2O_2 , $\text{Pd@Co}_3\text{O}_4/\text{GCE}$ rapidly attains the maximum steady-state current within 3 s. The amperometric response is linear to H_2O_2 concentration up to 3.3 mM, with a detection limit of 0.1 μM ($S/N = 3$), as shown in Figure 3E, and detection sensitivity is as high as 0.24 $\text{mA cm}^{-2} \text{mM}^{-1}$ (Figure S8, Supporting Information). The sensor performances of linear range, detection limit, and sensitivity in this work are superior to those of the previously reported electrochemical sensors based on noble metal catalysts and/or enzyme electrode for the determination of H_2O_2 .^{43–50} The reproducibility was explored by measuring 0.1 mM H_2O_2 using six different modified electrodes. The results show that the amperometric responses of six different $\text{Pd@Co}_3\text{O}_4/\text{GCEs}$

provide a relative standard deviation (RSD) value less than 5% (Figure 3F). More importantly, the amperometric responses of $\text{Pd@Co}_3\text{O}_4/\text{GCE}$ can maintain over 90% of its initial value after 50 times repeated testing or storage for more than 4 months, indicating the good reproducibility and stability of the proposed nanohybrid electrode for the detection of H_2O_2 .

Considering that the selectivity is an important parameter of biosensors, the potential interferences of several foreign species in real cells, including ascorbic acid (AA), uric acid (UA), dopamine (DA), glutamic acid, glycine, glutathione, Na^+ , K^+ , Mg^{2+} , Ca^{2+} , H^+ , and Cl^- , have been investigated. Our results showed that the changes of amperometric responses of 1.0 mM H_2O_2 were less than 10% upon the addition of 1.0 mM AA, UA, DA, glutamic acid, glycine, and glutathione and 5.0 mM Na^+ , K^+ , Mg^{2+} , Ca^{2+} , H^+ , and Cl^- (Table S1, Supporting Information), which indicates that the existence of these foreign species did not cause significant interference on the amperometric detection of H_2O_2 , and the nanohybrid electrode possesses high selectivity toward the electrochemical detection of H_2O_2 .

The high electrocatalytic activity and superior sensing performance of $\text{Pd@Co}_3\text{O}_4/\text{GCE}$ toward H_2O_2 can be attributed to two factors: On one hand, for the preparation of the $\text{Pd@Co}_3\text{O}_4$ precursor, i.e., $\text{SiO}_2@\text{PDA}@\text{Pd}$ sphere, the K_2PdCl_4 was in situ reduced by catechol groups of PDA on a SiO_2 sphere under the mild condition, leading to the formation of ultrafine and low-load Pd NPs, which are well-distributed on the surface of the SiO_2 sphere. For electrocatalytic application, the active metal NPs with fine particle size and uniform size distribution are anticipated to possess a high catalytic activity. On the other hand, for a $\text{Pd@Co}_3\text{O}_4$ hollow sphere, the microporous Co_3O_4 shells provide a large surface area for catalytic reaction as well as promote diffusion and mass transfer of reactants. And the Co_3O_4 shells can also prevent the encapsulated Pd NPs from aggregation or leaching by the shell protection, which endow the good stability of the resultant nanohybrid electrode.

Owing to the optimal performances of high sensitivity, wide linear range, and good reproducibility, the proposed $\text{Pd@Co}_3\text{O}_4$ based biosensor was used for in vitro tracking of H_2O_2 secretion by living cells. In this work, three types of living human cells, i.e., human cervical carcinoma HeLa cells, RSC96 Schwann cells, and human umbilical vein endothelial cells (HUVEC), were chosen for real-time electrochemical determination of H_2O_2 released upon being stimulated. The HeLa cell

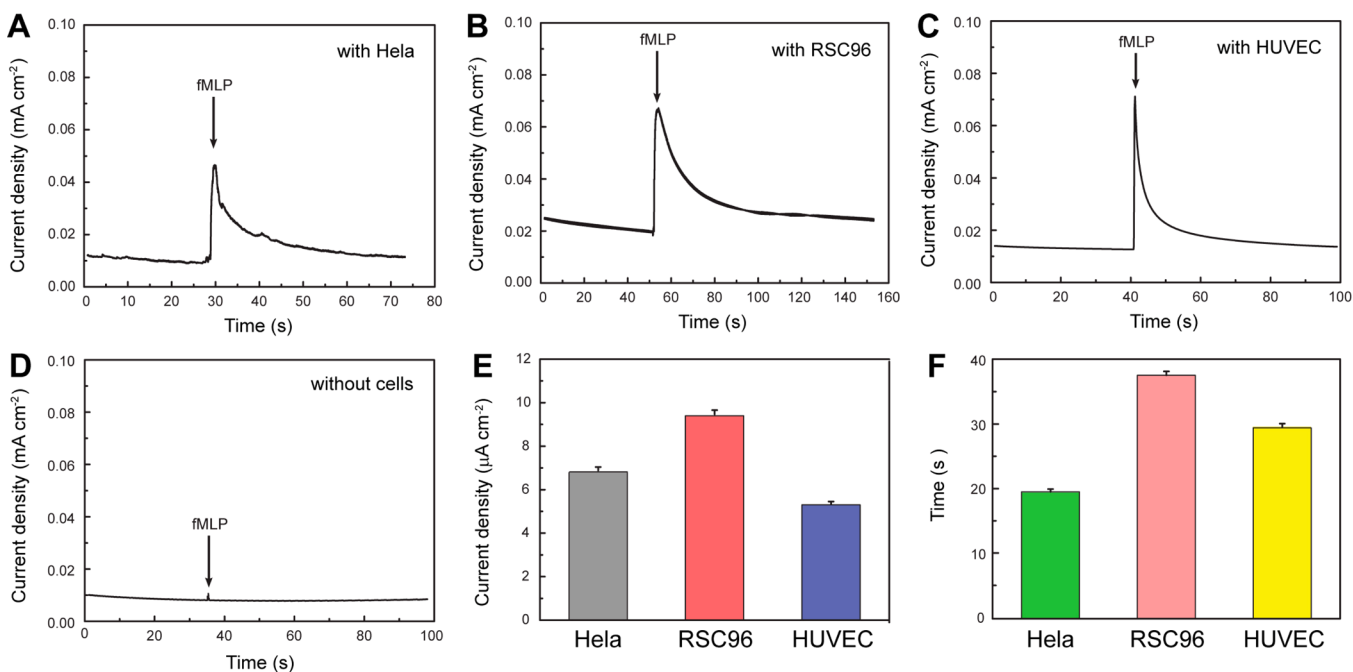


Figure 5. Amperometric responses of Pd@Co₃O₄/GCE in 0.1 M PBS (pH 7.4) with the addition of 10 μL fMLP (0.1 mM) in the presence of (A) HeLa cell, (B) RSC96 Schwann cells, and (C) HUVEC cells and (D) in the absence of cells. (E) The increased amperometric responses of three types of living cells upon the addition of 10 μL of fMLP (0.1 mM). (F) The response times to reach the plateau for three types of living cells.

line that derived from human cervical cancer cells is of great significance in cervical cancer diagnosis and treatment.⁵¹ The RSC96 cell line that derived from the glia of the peripheral nervous system is commonly used in nerve biologic research.⁵² And the HUVEC line that derived from the umbilical cord is used as a laboratory model system to study the function and pathology of endothelial cells.⁵³ Considering the different derivation and function of these three types of living human cells, the systematic study of their responses under the stimulation will be of great significance in physiological and pathological investigations. Figure 4 shows the bright-field and dark-field microscope images of HeLa cells, RSC96 cells, and HUVEC cells after being incubated with the Pd@Co₃O₄/GCE for over 2 h, which demonstrates that these three cells still maintained their activity, and did not show any changes in cell morphology and viability, indicating the good biocompatibility of Pd@Co₃O₄/GCE.

The live cells of 80% confluency are induced to release H₂O₂ by injecting fMLP, which is one of the synthetic *N*-formyl methionyl peptides that can simulate the secretion of several effector molecules including interleukin 1, lysosomal enzymes, tumor necrosis factor, nitric oxide, and reactive oxygen intermediates (ROI) by living cells.⁵⁴ Figure 5A–C depicts the amperometric response changes of Pd@Co₃O₄/GCE located near the living cell lines in a culture medium consisting of Dulbecco's modified Eagle medium (DMEM) at 37 °C. After injection of 10 μL of fMLP (0.1 mM), a significant increased current is observed for all the living cells at an applied potential of 0.6 V, followed by a gradual decrease of current, which finally reaches a plateau. However, the control wells containing no living cells do not generate any signal response to the addition of fMLP (Figure 5D). These results together suggest that the generated increase of amperometric responses is ascribed to the electrochemical reduction of H₂O₂ released by living cells upon being stimulated by fMLP. Moreover, the increased current densities and the times to reach the plateau for these three

types of living cells are investigated in detail. As shown in Figure 5E and 5F, upon the addition of an equal amount of fMLP, the increased amperometric current densities for HeLa cells, RSC96 cells, and HUVEC cells are 6.9, 9.2, and 5.3 μA cm⁻², respectively. And the response times to reach the plateau for these cells also increase in the order of RSC96 cells > HUVEC cells > HeLa cells. These observations substantially demonstrate that the proposed biosensor based on Pd@Co₃O₄/GCE establishes a simple, fast, and reliable method for the quantitative determination of H₂O₂ secreted by different living cells.

4. CONCLUSIONS

In summary, we have demonstrated the design and synthesis of a new type of high-performance nanocatalyst, i.e., microporous Pd@Co₃O₄ hollow nanosphere, and explored its practical application as an electrode material in an electrochemical sensor for in situ molecular detection of living cells. Owing to the synergy effect between the permeable microporous Co₃O₄ shell and the active Pd NPs that encapsulated in it, the resultant Pd@Co₃O₄ based biosensor shows significantly improved electrochemical sensing performance toward H₂O₂, even when the content of active Pd NPs in a Pd@Co₃O₄ hollow nanosphere is as low as 1.14 wt %. These enable it to be used for real-time tracking of the secretion of H₂O₂ in living human cells. The modular nature of this approach coupled with recent progress in core-shell hollow nanosphere synthesis and electrocatalysis opens new possibilities to systematically study the dependence of catalytic performance on the structural parameters and chemical compositions of the core-shell hollow nanosphere catalysts. Therefore, it is envisioned that the fabrication of Pd@Co₃O₄ provides a blueprint for designing various nanocatalysts encapsulated in microporous hollow sphere structures, which will be quite promising for a wide range of applications in catalysis, drug/gene delivery, and energy storage systems.

■ ASSOCIATED CONTENT

5 Supporting Information

SEM images of C@Pd hollow spheres and Pd@Co₃O₄ hollow spheres, TEM images of SiO₂ spheres and Pd@Co₃O₄ hollow spheres, EDX image of C@Pd hollow spheres, XRD pattern of Pd@Co₃O₄ hollow nanospheres, the relationship curves between the amperometric current responses of Pd@Co₃O₄/GCE toward 2.0 mM H₂O₂ and mass ratio of K₂PdCl₄ to SiO₂@PDA, calibration curves of the amperometric response of Pd@Co₃O₄/GCE to successive addition of H₂O₂ in stirring PBS buffer (pH 7.4), and the table of influence of foreign species on the determination of 1.0 mM H₂O₂. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) Luo, Y. P.; Liu, H. Q.; Rui, Q. Y.; Tian, Y. Detection of Extracellular H₂O₂ Released from Human Liver Cancer Cells Based on TiO₂ Nanoneedles with Enhanced Electron Transfer of Cytochrome c. *Anal. Chem.* **2009**, *81*, 3035–3041.
- (2) Luo, Y. P.; Tian, Y.; Rui, Q. Electrochemical Assay of Superoxide Based on Biomimetic Enzyme at Highly Conductive TiO₂ Nanoneedles: From Principle to Applications in Living Cells. *Chem. Commun.* **2009**, *21*, 3014–3016.
- (3) Miller, E. W.; Dickinson, B. C.; Chang, C. J. Aquaporin-3 Mediates Hydrogen Peroxide Uptake to Regulate Downstream Intracellular Signaling. *Proc. Natl. Acad. Sci., India* **2010**, *7*, 15681–15686.
- (4) Yuan, L.; Lin, W.; Xie, Y.; Chen, B.; Zhu, S. Single Fluorescent Probe Responds to H₂O₂, NO, and H₂O₂/NO with Three Different Sets of Fluorescence Signals. *J. Am. Chem. Soc.* **2012**, *134*, 1305–1315.
- (5) Xu, J.; Li, Q.; Yue, Y.; Guo, Y.; Shao, S. J. A Water-soluble BODIPY Derivative as a Highly Selective “Turn-On” Fluorescent Sensor for H₂O₂ Sensing in Vivo. *Biosens. Bioelectron.* **2014**, *56*, 58–63.
- (6) Luo, W.; Abbas, M. E.; Zhu, L.; Deng, K.; Tang, H. Rapid Quantitative Determination of Hydrogen Peroxide by Oxidation Decolorization of Methyl Orange Using a Fenton Reaction System. *Anal. Chim. Acta* **2008**, *629*, 1–5.
- (7) Effkemann, S.; Pinkernell, U.; Neumuller, R.; Schwan, F.; Engelhardt, H.; Karst, U. Liquid Chromatographic Simultaneous Determination of Peroxycarboxylic Acids Using Postcolumn Derivatization. *Anal. Chem.* **1998**, *70*, 3857–3862.
- (8) Šljukić, B.; Banks, C. E.; Compton, R. G. Iron Oxide Particles Are the Active Sites for Hydrogen Peroxide Sensing at Multiwalled Carbon Nanotube Modified Electrodes. *Nano Lett.* **2006**, *6*, 1556–1558.
- (9) Li, J. W.; Liu, L. H.; Xiao, F.; Gui, Z.; Yan, R.; Zhao, F. Q.; Hu, L.; Zeng, B. Z. Direct Electron Transfer and Electrocatalysis of Horseradish Peroxidase Immobilized in Gemini Surfactant–Ionic Liquid Composite Film on Glassy Carbon Electrode. *J. Electroanal. Chem.* **2008**, *613*, 51–57.

- (10) Li, P.; Ding, Y.; Lu, Z. Y.; Chen, Y.; Zhou, Y. M.; Tang, Y. W.; Cai, C. X.; Lu, T. H. An ultrasensitive Iron(III)-complex Based Hydrogen Peroxide Electrochemical Sensor Based on A Non-electrocatalytic Mechanism. *Anal. Chim. Acta* **2013**, *786*, 34–38.

- (11) Lin, L.; Yan, J.; Li, J. H. Small-Molecule Triggered Cascade Enzymatic Catalysis in Hour-Glass Shaped Nanochannel Reactor for Glucose Monitoring. *Anal. Chem.* **2014**, *86*, 10546–10551.

- (12) Lippert, A. R.; DeBittner, G. C. V.; Chang, C. J. Boronate Oxidation as a Bioorthogonal Reaction Approach for Studying the Chemistry of Hydrogen Peroxide in Living Systems. *Acc. Chem. Res.* **2011**, *44*, 793–804.

- (13) Zhou, L. Z.; Kuai, L.; Li, W. Z.; Geng, B. Y. Ion-Exchange Route to Au-Cu_xOS Yolk-Shell Nanostructures with Porous Shells and Their Ultrasensitive H₂O₂ Detection. *ACS Appl. Mater. Interfaces* **2012**, *4*, 6463–6467.

- (14) Mazumder, V.; Sun, S. H. Oleylamine-Mediated Synthesis of Pd Nanoparticles for Catalytic Formic Acid Oxidation. *J. Am. Chem. Soc.* **2009**, *131*, 4588–4589.

- (15) Huang, X. Q.; Tang, S. H.; Zhang, H. H.; Zhou, Z. Y.; Zheng, N. F. Controlled Formation of Concave Tetrahedral/Trigonal Bipyramidal Palladium Nanocrystals. *J. Am. Chem. Soc.* **2009**, *131*, 13916–13917.

- (16) Lee, H. J.; Habas, S. E.; Somorjai, G. A.; Yang, P. D. Localized Pd Overgrowth on Cubic Pt Nanocrystals for Enhanced Electrocatalytic Oxidation of Formic Acid. *J. Am. Chem. Soc.* **2008**, *130*, 5406–5407.

- (17) Han, M.; Liu, S. L.; Bao, J. C.; Dai, Z. H. Pd Nanoparticle Assemblies—As the Substitute of HRP, in Their Biosensing Applications for H₂O₂ and Glucose. *Biosens. Bioelectron.* **2012**, *31*, 151–156.

- (18) Joo, S. H.; Park, J. Y.; Tsung, C. K.; Yamada, Y.; Yang, P.; Somorjai, G. A. Thermally Stable Pt/mesoporous Silica Core-shell Nanocatalysts for High-temperature Reactions. *Nat. Mater.* **2009**, *8*, 126–131.

- (19) Lopez-Sanchez, J. A.; Dimitratos, N.; Hammond, C.; Brett, G. L.; Kesavan, L.; White, S.; Miedzkiak, P.; Tiruvalam, R.; Jenkins, R. L.; Carley, A. F.; Knight, D.; Kiely, C. J.; Hutchings, G. J. Facile Removal of Stabilizer-ligands from Supported Gold Nanoparticles. *Nat. Chem.* **2011**, *3*, 551–556.

- (20) Yoon, K.; Yang, Y.; Lu, P.; Wan, D. H.; Peng, H. C.; Masias, K. S.; Fanson, P. T.; Campbell, C. T.; Xia, Y. N. A Highly Reactive and Sinter-Resistant Catalytic System Based on Platinum Nanoparticles Embedded in the Inner Surfaces of CeO₂ Hollow Fibers. *Angew. Chem., Int. Ed.* **2012**, *51*, 9543–9546.

- (21) Bo, X. J.; Bai, J.; Ju, J.; Guo, L. P. A Sensitive Amperometric Sensor for Hydrazine and Hydrogen Peroxide Based on Palladium Nanoparticles/onion-like Mesoporous Carbon Vesicle. *Anal. Chim. Acta* **2010**, *675*, 29–35.

- (22) Zhang, F.; Yuan, C. Z.; Zhu, J. J.; Wang, J.; Zhang, X. G.; Lou, X. W. Flexible Films Derived from Electrospun Carbon Nanofibers Incorporated with Co₃O₄ Hollow Nanoparticles as Self-Supported Electrodes for Electrochemical Capacitors. *Adv. Funct. Mater.* **2013**, *23*, 3909–3915.

- (23) Lou, X. W. D.; Yuan, C.; Rhoades, E.; Zhang, Q.; Archer, L. A. Encapsulation and Ostwald Ripening of Au and Au–Cl Complex Nanostructures in Silica Shells. *Adv. Funct. Mater.* **2006**, *16*, 1679–1684.

- (24) Zhang, G. Q.; Lou, X. W. D. General Synthesis of Multi-Shelled Mixed Metal Oxide Hollow Spheres with Superior Lithium Storage Properties. *Angew. Chem., Int. Ed.* **2014**, *53*, 9041–9044.

- (25) Zhang, G. Q.; Lou, X. W. D. General Synthesis of Multi-Shelled Mixed Metal Oxide Hollow Spheres with Superior Lithium Storage Properties. *Angew. Chem., Int. Ed.* **2014**, *53*, 9041–9044.

- (26) Jiao, Q.; Fu, Z. M.; You, C.; Zhao, Y.; Li, H. S. Preparation of Hollow Co₃O₄ Microspheres and Their Ethanol Sensing Properties. *Inorg. Chem.* **2012**, *51*, 11513–11520.

- (27) Wang, J. Y.; Yang, N. L.; Tang, H. J.; Dong, Z. H.; Jin, Q.; Yang, M.; Kisailus, D.; Zhao, H. J.; Tang, Z. Y.; Wang, D. Accurate Control of Multishelled Co₃O₄ Hollow Microspheres as High-Performance

Anode Materials in Lithium-Ion Batteries. *Angew. Chem., Int. Ed.* **2013**, *125*, 6545–6548.

(28) Liu, J.; Qiao, S. Z.; Chen, J. S.; Lou, X. W. D.; Xing, X.; Lu, G. Q. M. Yolk/shell Nanoparticles: New Platforms for Nanoreactors, Drug Delivery and Lithium-ion Batteries. *Chem. Commun.* **2011**, *47*, 12578–12591.

(29) Lou, X. W. D.; Archer, L. A.; Yang, Z. C. Hollow Micro-/Nanostructures: Synthesis and Applications. *Adv. Mater.* **2008**, *20*, 3987–4019.

(30) Liu, W.; Liu, J.; Chen, K. F.; Ji, S. M.; Wan, Y. L.; Zhou, Y. C.; Xue, D. F.; Hodgson, P.; Li, Y. C. Enhancing the Electrochemical Performance of the LiMn_2O_4 Hollow Microsphere Cathode with a $\text{LiNi}_{0.5}\text{Mn}_{1.5}\text{O}_4$ Coated Layer. *Chem.—Eur. J.* **2014**, *20*, 824–830.

(31) Liu, J.; Liu, W.; Ji, S. M.; Wan, Y. L.; Gu, M. Z.; Yin, H. Q.; Zhou, Y. C. Iron Fluoride Hollow Porous Microspheres: Facile Solution-Phase Synthesis and Their Application for Li-Ion Battery Cathodes. *Chem.—Eur. J.* **2014**, *20*, 5815–5820.

(32) Liu, J.; Liu, W.; Chen, K. F.; Ji, S. M.; Zhou, Y. C.; Wan, Y. L.; Xue, D. F.; Hodgson, P.; Li, Y. C. Facile Synthesis of Transition-Metal Oxide Nanocrystals Embedded in Hollow Carbon Microspheres for High-Rate Lithium-Ion-Battery Anodes. *Chem.—Eur. J.* **2013**, *19*, 9811–9816.

(33) Liu, J.; Xia, H.; Xue, D. F.; Lu, L. Double-shelled Nanocapsules of V_2O_5 -Based Composites as High-performance Anode and Cathode Materials for Li Ion Batteries. *J. Am. Chem. Soc.* **2009**, *131*, 12086–12087.

(34) Zhang, N.; Xu, Y. J. Aggregation- and Leaching-Resistant, Reusable, and Multifunctional $\text{Pd}@ \text{CeO}_2$ as a Robust Nanocatalyst Achieved by a Hollow Core–Shell Strategy. *Chem. Mater.* **2013**, *25*, 1979–1988.

(35) Lee, H.; Dellatore, S. M.; Miller, W. M.; Messersmith, P. B. Mussel-Inspired Surface Chemistry for Multifunctional Coatings. *Science* **2007**, *318*, 426–430.

(36) Guo, L. Q.; Liu, Q.; Li, G. Q.; Shi, J. B.; Liu, J. Y.; Wang, T.; Jiang, G. B. A Mussel-Inspired Polydopamine Coating as a Versatile Platform for the In Situ Synthesis of Graphene-based Nanocomposites. *Nanoscale* **2012**, *4*, 5864–5867.

(37) Zhang, Z. Y.; Xiao, F.; Xi, J. B.; Sun, T.; Xiao, S.; Wang, H. R.; Wang, S.; Liu, Y. Q. Encapsulating Pd Nanoparticles in Double-Shell Graphene@Carbon Hollow Spheres for Excellent Chemical Catalytic Property. *Sci. Rep.* **2014**, *4*, 4053.

(38) Liu, Y.; Ai, K.; Lu, L. Polydopamine and Its Derivative Materials: Synthesis and Promising Applications in Energy, Environmental, and Biomedical Fields. *Chem. Rev.* **2014**, *114*, 5057–5115.

(39) Ball, V.; Nguyen, I.; Haupt, M.; Oehr, C.; Arnoult, C.; Toniazzo, V.; Ruch, D. The Reduction of Ag^+ in Metallic Silver on Pseudomelanin Films Allows For Antibacterial Activity But Does Not Imply Unpaired Electrons. *J. Colloid Interface Sci.* **2011**, *364*, 359–365.

(40) Xie, X. W.; Li, Y. Z.; Liu, Q.; Haruta, M.; Shen, W. J. Low-temperature Oxidation of CO Catalysed by Co_3O_4 Nanorods. *Nature* **2009**, *458*, 746–749.

(41) Ye, J. Y.; Liu, C. J. $\text{Cu}_3(\text{BTC})_2$: CO Oxidation Over MOF Based Catalysts. *Chem. Commun.* **2011**, *47*, 2167–2169.

(42) Gao, X. Q.; Li, F. M.; Li, Y. M.; Li, S. N.; Chen, Y.; Lee, J. M. A Surfactant-free Strategy for Synthesizing Reduced Graphene Oxide Supported Palladium Nanoparticles with Enhanced Electrocatalytic Performance towards Formic Acid Oxidation. *J. Power Sources* **2015**, *280*, 491–498.

(43) Xu, F. G.; Sun, Y. J.; Zhang, Y.; Shi, Y.; Wen, Z. W.; Li, Z. Graphene–Pt Nanocomposite for Nonenzymatic Detection of Hydrogen Peroxide with Enhanced Sensitivity. *Electrochem. Commun.* **2011**, *13*, 1131–1134.

(44) Guascito, M. R.; Chirizzi, D.; Malitesta, C.; Mazzotta, E.; Siciliano, M.; Siciliano, T.; Tepore, A.; Turco, A. Low-potential Sensitive H_2O_2 Detection Based on Composite Micro tubular Te Adsorbed on Platinum Electrode. *Biosens. Bioelectron.* **2011**, *26*, 3562–3569.

(45) Xiao, F.; Song, J. B.; Gao, H. C.; Zan, X. L.; Xu, R.; Duan, H. W. Coating Graphene Paper with 2D-assembly of Electrocatalytic Nanoparticles: A Modular Approach Toward High-performance Flexible Electrodes. *ACS Nano* **2012**, *6*, 100–110.

(46) Chen, K. J.; Pillai, K. C.; Rick, J.; Pan, C. J.; Wang, S. H.; Liu, C. C.; Hwang, B. J. Bimetallic PtM (M = Pd, Ir) Nanoparticle Decorated Multi-walled Carbon Nanotube Enzyme-free, Mediator-less Amperometric Sensor for H_2O_2 . *Biosens. Bioelectron.* **2012**, *33*, 120–127.

(47) Han, Y.; Zheng, J.; Dong, S. A Novel Nonenzymatic Hydrogen Peroxide Sensor Based on Ag– MnO_2 –MWCNTs Nanocomposites. *Electrochim. Acta* **2013**, *90*, 35–43.

(48) Ji, S.; Guo, Q.; Yue, Q.; Wang, L.; Wang, H.; Zhao, J.; Dong, R.; Liu, J.; Jia, J. Controlled Synthesis of Pt Nanoparticles Array Through Electroreduction of Cisplatin Bound at Nucleobases Terminated Surface and Application into H_2O_2 Sensing. *Biosens. Bioelectron.* **2011**, *26*, 2067–2073.

(49) Miao, Y. E.; He, S.; Zhong, Y.; Yang, Z.; Tjiu, W. W.; Liu, T. A Novel Hydrogen Peroxide Sensor Based on Ag/ SnO_2 Composite Nanotubes by Electrospinning. *Electrochim. Acta* **2013**, *99*, 117–123.

(50) Suárez, G.; Santschi, C.; Martin, O. J. F.; Slaveykova, V. Biosensor Based on Chemically-designed Anchorable Cytochrome *c* for the Detection of H_2O_2 Released by Aquatic Cells. *Biosens. Bioelectron.* **2013**, *42*, 385–390.

(51) Rahbari, R.; Sheahan, T.; Modes, V.; Collier, P.; Macfarlane, C.; Badge, R. M. A novel L1 Retrotransposon Marker for HeLa Cell Line Identification. *Biotechniques* **2009**, *46*, 277–284.

(52) Ji, Y.; Shen, M.; Wang, X.; Zhang, S.; Yu, S.; Chen, G.; Gu, X.; Ding, F. Comparative Proteomic Analysis of Primary Schwann Cells And a Spontaneously Immortalized Schwann Cell Line RSC 96: A Comprehensive Overview With a Focus on Cell Adhesion and Migration Related Proteins. *J. Proteome Res.* **2012**, *11*, 3186–3198.

(53) Liu, L. S.; Wei, D. H.; Tang, C. K.; Wang, G. X.; Zhang, S. C.; Yin, W. D.; Yang, Y. Z.; Legrand, A. P.; Guidoin, R. A HUVEC Line with a Stable Expression of the VEGF121 Gene to Achieve Complete Endothelialization of Blood Conduits. *Artif. Cells, Blood Substitutes, Immobilization Biotechnol.* **2007**, *35*, 319–331.

(54) Ijzermans, J. N.; Marquet, R. L. Interferon-gamma: A Review. *Immunobiology* **1989**, *179*, 456–473.